

METHODOLOGY

Open Access



3D-printed mouthpiece adapter for sampling exhaled breath in medical applications

Y Lan Pham^{1,2}, Jonathan Beauchamp¹, Alexander Clement³, Felix Wiegandt³ and Olaf Holz^{3,4*}

Abstract

The growing use of 3D printing in the biomedical sciences demonstrates its utility for a wide range of research and healthcare applications, including its potential implementation in the discipline of breath analysis to overcome current limitations and substantial costs of commercial breath sampling interfaces. This technical note reports on the design and construction of a 3D-printed mouthpiece adapter for sampling exhaled breath using the commercial respiration collector for in-vitro analysis (ReCIVA) device. The paper presents the design and digital workflow transition of the adapter and its fabrication from three commercial resins (Surgical Guide, Tough v5, and BioMed Clear) using a Formlabs Form 3B stereolithography (SLA) printer. The use of the mouthpiece adapter in conjunction with a pulmonary function filter is appraised in comparison to the conventional commercial silicon facemask sampling interface. Besides its lower cost – investment cost of the printing equipment notwithstanding – the 3D-printed adapter has several benefits, including ensuring breath sampling via the mouth, reducing the likelihood of direct contact of the patient with the breath sampling tubes, and being autoclaveable to enable the repeated use of a single adapter, thereby reducing waste and associated environmental burden compared to current one-way disposable facemasks. The novel adapter for breath sampling presented in this technical note represents an additional field of application for 3D printing that further demonstrates its widespread applicability in biomedicine.

Keywords: Prototyping, Sampling Interface, Resin-printed Device, Stereolithography, Breath Analysis, Spirometry

Introduction

3D printing for healthcare applications has experienced a surge in interest in recent years due to advancements in the technical performances of printers, the emergence of suitable printing composite materials, and the widespread availability of low-cost printing devices [1, 2]. The rapid growth of 3D printing applications in the medical field is expected to revolutionise healthcare [3] and has been suggested to be of particular benefit in times of health crises when 3D printed materials can offer on-demand

alternatives to medical parts and equipment that may be in short supply [4–7]. Prevalent biomedical applications include customised prosthetics [8, 9], implants [10], tissue and organ fabrication [11, 12], or anatomical models for medical training purposes [13–15], amongst others. Since their first reported use in medicine in the early 2000s [16, 17], 3D printers have evolved to become a reliable engineering tool in this field, providing the freedom to produce bespoke medical products and fittings with enhanced and cost-effective productivity [18].

A hitherto niche area within the expansive field of medical research is breath analysis, which has emerged in recent years as a promising approach to non-invasive disease diagnostics [19]. Exhaled breath contains a rich mixture of volatile chemical compounds [20], and the

*Correspondence: olaf.holz@item.fraunhofer.de

⁴ Member of the German Centre of Lung Research DZL (BREATH), Hannover, Germany

Full list of author information is available at the end of the article



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

basic premise of breath analysis is to examine these constituents as a means to seek (early) indications of illness or infection. The benefits of breath analysis over conventional diagnostics using blood are its non-invasive (painless) sampling combined with rapid detection, offering the potential for widespread implementation in the areas of clinical diagnosis, routine screening, and therapeutic monitoring [21, 22].

In breath research, exhaled breath can be sampled for either on-line or off-line analysis. The former involves the direct connection of a breath sampling interface to the analyser to allow for immediate analysis of exhaled breath gas [23], whereas the latter involves the collection of a breath sample in a storage medium, which is then transferred to the analytical platform for subsequent analysis [21]; the respective benefits and drawbacks of these two approaches are discussed in the scientific literature [24, 25]. The most prevalent approach to sampling exhaled breath for breath gas analysis is the off-line method.

Due to the broad nature of breath research, whereby a wide variety of analytical technologies are used to target an extensive range of compounds for a diverse assortment of illnesses, a standardised approach to sampling for off-line analysis does not exist [26]. Off-line sampling methods include the collection of breath into inflatable bags, or its collection and pre-concentration onto adsorbent materials, allowing for trace level detection of breath-borne volatiles [27]. The most common approach utilises thermal desorption (TD) tubes – i.e., hollow tubes containing an adsorbent material with a high affinity to organic compounds – for sample collection, which are then interfaced with a gas chromatography-mass spectrometry (GC-MS) analyser, whereby the tubes are heated to liberate the trapped compounds into the analyser, i.e., thermal desorption, to enable their subsequent detection [28].

Sampling systems to collect breath volatiles on TD tubes are often custom built by individual research groups, but commercial devices exist, such as the Respiration Collector for In-Vitro Analysis (ReCIVA) breath sampler (Owlstone Medical Ltd., Cambridge, UK), which captures end-tidal breath directly onto TD tubes using a capnography-driven sampling protocol and associated control software [29]. The breath collector is conventionally equipped with a silicon facemask – similar to an oxygen mask used for ventilation in the clinical setting – that offers a snug fit over the mouth and nose of the participant and allows for rebreathing and sample collection [29–32]. Four holes at the bottom of the mask allow for insertion of the TD tubes, which protrude into the inside of the mask, located directly in-line with the wearer's breathing flow. An integrated sterility filter at the downstream end of the mask ensures that the ReCIVA device is

shielded from exposure to aerosols in the exhalation flow, thereby reducing the risk of cross-contamination when using the same device successively for different patients.

Despite some benefits of convenience, this facemask has several drawbacks. Foremost, its design does not permit the control of breathing solely via the mouth, but rather allows for mixed nasal-oral breathing; in breath analysis, the breathing route can directly impact the type and abundance of volatiles, since some compounds have production sites in the nasal cavity, whereas others originate from the oral cavity [33–35], thus a consistent exhalation route is imperative. Another limitation of the mask is the very close proximity of the TD tubes to the wearer's face (mouth and nose), resulting in a high likelihood of direct physical contact to the non-sterile tubes (depending on anatomy) with an associated risk of infection to the potentially vulnerable patient, as well as a propensity to contaminate the tubes with aerosols present in exhaled breath (see Figure S1). Finally, the masks exhibit a short shelf-life of sterility and single-use performance, which are associated with considerable cost and waste for large cohort studies.

To address these shortcomings, an alternative mouth-piece adapter for connection to the ReCIVA device was designed for 3D stereolithography (SLA) printing to replace the facemask as a cost-effective and reusable alternative that ensures controlled exhalation through the mouth and a lesser likelihood of direct contact between the patient and the sampling tubes, thus reducing the risk of infection and cross-contamination between patients.

This technical note presents details of the design and construction of this inexpensive 3D-printed interface, with a view to making it available for wider use and potential adaptation and adoption by the breath research community in combination with the ReCIVA breath collection device as a viable alternative to the silicon facemask.

Methods

A routine workflow for model design and 3D-printing was undertaken, namely to establish an *in silico* three-dimensional model of the object for subsequent 3D printing into its physical representation. In a first step, a 3D prototype model was created using Autodesk Inventor Professional computer-aided design (CAD) software (Inventor 2015 Build: 159, Release 2015 RTM., Autodesk Inc., CA, USA). The 3D model was exported to Standard Tessellation Language (STL) format, which contains mesh coordinates of 3D models (depicted in Fig. 1). Next, the Formlabs PreForm software (v. 3.19.1., Formlabs Inc., Somerville, MA, USA) was used to convert the STL file into a 3D printable format (.form), which includes printing information, such as layer thickness, printing

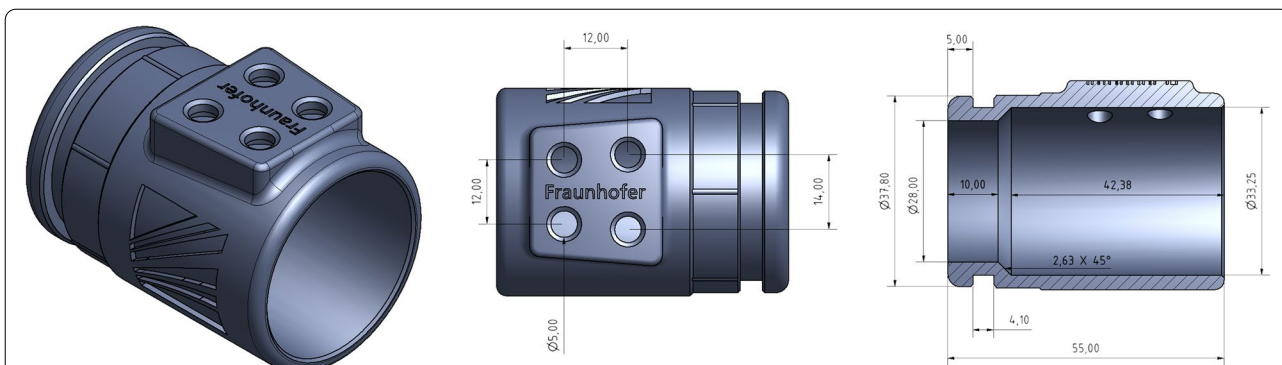


Fig. 1 Model of the breath sampling adapter, viewed from three perspectives. Left: oblique view, showing the connection to the sampling device (facing rear-left) and the underside of the adapter (top/centre); centre: bottom view, showing the underside of the adapter and the connection orifices for the four adsorbent tubes in trapezoid formation; and right: cross-sectional planar view, showing the side of the adapter (the adapter connects to the breath sampling device on the left-hand (rear) end; a pulmonary function filter is attached at the right-hand (front) end); the base of the adapter, containing the connection orifices for the adsorption tubes, is depicted at the top. All dimensions are in mm

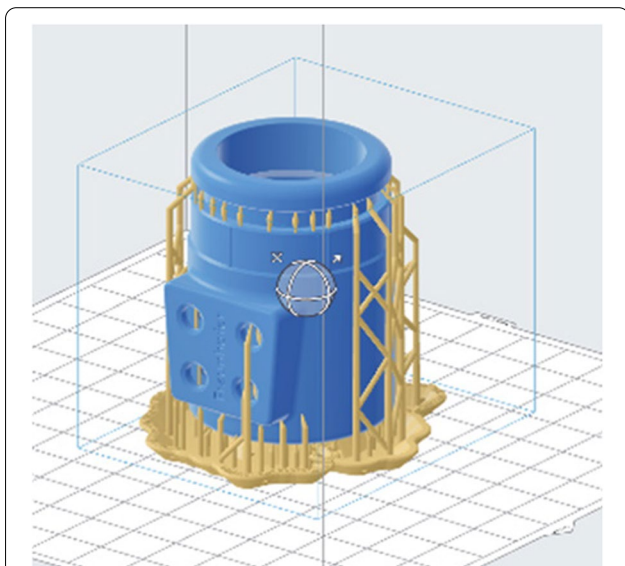


Fig. 2 Model of the adapter (blue) for 3D printing in an upright position, displaying the vertical support structures (ochre colour)

material, orientation, and support structures (see Figs. 2 and 3) in order to facilitate printing. Finally, the ensuing files were exported for the individual resin materials and transferred to the printer for 3D printing.

CAD model and 3D-printing of the adapter

The initial concept and development of the breath sampling adapter was motivated by a need to overcome the limitations of the current commercial silicon facemasks. This modification centres on an adapter that can be mounted directly to the ReCIVA breath sampling device and enables the simultaneous collection of volatile constituent of breath onto up to four independent TD tubes by exhaling through the hollow cylindrical adapter. The adapter can be paired with a commercially available pulmonary function filter to reduce the risk of cross-infection between participants (see Fig. 4) and the adapter itself can be treated for reuse, e.g., by autoclaving.

The core design of the adapter is a cylindrical tube of 55 mm length and 39 mm outer diameter. Its inner diameter



Fig. 3 Final adapter models pictured with (left) and without (right) support structures, printed from Tough v5 (blue), Surgical Guide (orange) and BioMed Clear (translucent) resins



Fig. 4 Left: Assembled breath sampling device comprising the ReCIVA breath sampler (right), a 3D-printed mouthpiece adapter (an adapter made from BioMed Clear resin is depicted; centre), a disposable pulmonary function filter (left), and thermal desorption tubes to sample breath (bottom-centre). The nose clip depicted is used as an adjunct during breath sampling to ensure mouth breathing. Right: Close-up of the underside of the adapter with three inserted thermal desorption tubes and one open tube orifice fitted with an O-ring

for most of its length is 33 mm, but with an inner lip of 28 mm at the downstream end where it connects to the sampling device, allowing it to click firmly into place (see Fig. 1). A trapezoid-shaped protrusion along the axial direction and located at the back half of the adapter allows for connection of the TD tubes. To enable their connection to the adapter, the protrusion houses four orifices of 5.0 mm diameter – subsequently enlarged post-printing to 6.3 mm (see later) – that are positioned in a slight trapezoid configuration. These holes are furnished with sunken rims to allow for the insertion of Viton O-rings (6.3 × 2.0 mm inner diameter × thickness) to provide a firm attachment of standard size TD tubes (0.25" × 3.50" outer diameter × length), as well as to ensure isolation of the tubes from ambient air exterior to the adapter (see Fig. 4). The aforementioned tapered rim at the rear end of the adapter allows for its firm connection to the ReCIVA device.

All models were printed using a Form 3B printer (or Form 2 printer for earlier prototypes; not depicted here) (Formlabs Inc.) and a selection of different resins, namely Tough v5, Surgical Guide and BioMed Clear (Formlabs Inc.). The use of three different resins was to explore their different properties and related suitability for breath sampling purposes. Printing proceeded using a 100 µm layer thickness build at a pre-programmed printing temperature of 35 °C. The support structures were created automatically by the printer. After printing, the uncured models underwent post-processing. This comprised: washing (whilst still attached to the print bed) for 20 min in >95% isopropyl alcohol (IPA) in a washing station (Form Wash, Formlabs Inc.) to ensure removal of any residual uncured resin; physical detachment from the print bed; post-curing under ultraviolet (UV) light

in a cure station (Form Cure, Formlabs Inc.) in accordance with the manufacturer guidelines specific to each resin material; and finally, manual removal of the support structures using cutting pliers. Additionally, any residual protrusions or rough areas remaining on the printed surfaces after removal of the support structures were mechanically removed by abrasion (fine sandpaper) to achieve a smooth surface. The final, printed adapters for all three resins are depicted in Fig. 3, with and without supporting structures.

Adapter use for breath sampling

Before the newly printed – or, subsequently, used – adapters may be used for their intended purpose, they must be treated to ensure cleanliness and sterility. One approach to achieve this is sterilisation by autoclaving (steaming). This process was undertaken and examined for the printed adapters, in order to assess their resilience to autoclaving and their performance after this treatment. Autoclaving proceeded by wrapping an individual adapter in aluminium foil (after removal of the O-rings) and applying steam sterilisation at 121 °C for 30 min using a benchtop autoclave (HMC 300 MBF; HMC Europe GmbH, Tüßling, Germany), followed by a 30 min drying phase. After this treatment, the adapters may be stored under sterile conditions (in packaging pouches) until their use for breath sampling. Alternative sterilisation methods include treatment with ethylene oxide (ETO) or plasma sterilisation. Autoclaving was used in the present study following manufacturer recommendations of steam sterilisation for these resins.

To assemble the breath sampling device, O-rings are first placed in the four orifices of the adapter, then (up to) four TD tubes are inserted into these orifices, ensuring a

snug fit (see Fig. 4). The other ends of the tubes are then pushed down into the four hollow insets at the bottom of the ReCIVA device. The ReCIVA device allows for sampling onto any one or all of the four tubes; if exhaled breath is to be sampled on less than four tubes, empty tubes or dummy rods of identical dimensions to the tubes can be inserted in the unused positions to maintain structural rigidity. After the tubes have been attached in the described manner, the front end of the adapter can be fitted with a pulmonary function filter, after which the grooved (rear) end is connected to the socket of the ReCIVA device, as depicted in Fig. 4.

Results and Discussion

CAD adapter design and 3D-printed models

The choice of resins used to produce the prototype adapters described herein was made according to their expected suitability for the intended purpose, based on the material datasheet specifications of the manufacturer. Tough v5 resin (discontinued, but available at the time of writing in modified composition as Tough 2000) is described as exhibiting a particularly sturdy structure, thus promising long-term durability. By comparison, Surgical Guide resin is promoted as non-cytotoxic, non-irritative and non-sensitising according to the ISO 10993-1:2018 standard, thereby representing a particularly attractive material for the adapter because of its intended use with human subjects. Lastly, BioMed Clear resin has passed biocompatibility requirements of the ISO 10993-1:2018, ISO 7405:2018 and ISO 18562-1:2017 standards, thus surpasses the properties of Surgical Guide resin by purportedly exhibiting no emissions of particulates, volatile organic compounds, or hazardous water-soluble substances. This latter aspect is essential in applications with direct contact to human subjects, and especially so in breath analysis to avoid inhalation of compounds or particulate matter on the one hand, and sampling of compound emissions as potential confounders on the other. An appraisal of the emissions is beyond the scope of this technical note but is currently under investigation for future dissemination.

Support structures of the printed model had different touchpoint sizes that ranged from 0.3 mm to 0.7 mm, depending on the resin used for printing. Trial printing runs for different orientations of the model yielded an upright position to be the most favourable for printing due to the absence of support structures on the inner surface of the adapter; in addition to the associated practical benefit of avoiding mechanical removal of these support structures at an inconvenient and poorly accessible location within the cylinder, the ensuing rough surface caused by the support structures are expected to have a higher adsorption potential of volatile compounds, which

would be of detriment to the intended application of sampling breath volatiles. A drawback of this print orientation, however, was that the four orifices for insertion of the TD tubes were not uniformly round but rather slightly oval in shape. This was overcome by printing the orifices with a smaller diameter (5 mm), then using a bur to mechanically increase this diameter to accommodate the TD tubes (6.6 mm), thereby providing a firm attachment of the conventional 0.25" diameter TD tubes. (It might be noted that printing in horizontal orientation produces uniformly round holes, but presents the drawback described above in relation to the presence of supporting structures within the adapter tube; the vertical printing approach was deemed the most suitable to ensure precision for the adapter connection to the sampling system, and offered flexibility in achieving a snug fit for insertion of the four TD tubes.) In terms of yield, a standard 1 L resin tank can produce approximately 45 adapters, depending on the resin.

Comparison of 3D-printed adapter with commercial facemask

Conventionally, the ReCIVA breath sampler is used with a silicon facemask to sample exhaled breath, which is delivered by the supplier (Owlstone Medical) under sterile conditions. The facemask offers the advantage of flexibility of the silicon that adapts to the contours of the wearer's face, ensuring a snug fit. A further convenience of the facemask is its integrated sterility filter at the downstream end to reduce the risk of contamination of the ReCIVA device by the passage of exhaled breath during use. While the latter provides a degree of protection to the ReCIVA device that lessens the risk of cross-infection between patients, the close proximity of the wearer's nose and mouth to the sampling tubes poses a risk of infection to the wearer, especially when breath is sampled from immunocompromised patients; the TD tubes cannot be treated sufficiently to eliminate this risk without compromising their sampling performance. The mouth-piece adapter presented here minimises this risk through use of a pulmonary function filter between the patient and the adapter, thus direct contact of the patient's lips or nose with the TD tubes is prohibited. In practice, this configuration requires the additional use of a nose clip to ensure that the participant exhales exclusively through the mouth and inhales clean air provided by the CASPER Portable Air Supply (Owlstone Medical Ltd., Cambridge, UK) attached to the ReCIVA device, as is also routine configuration when using the facemask. However, the use of a nose clip can be considered an advantage, since sampling breath via the mouth is usually desirable in breath analysis, as discussed above.

In addition to the aforementioned limitations of the conventional facemask for sampling, the mask is relatively high in cost (ca. \$ 25) for a single-use, disposable item and exhibits a limited sterility period of two weeks after purchase (treatment), as stipulated by the supplier. This represents a hindrance for its widespread cost-effective implementation for breath tests in routine analysis or large cohort clinical studies, and leads to high waste and an associated environmental burden. By comparison, the novel 3D-printed adapter presented herein exhibits properties that overcome some of the limitations of the facemasks. Considering expense, the cost of producing a single adapter is between \$ 4-8, depending on the resin and based on current commercial prices (investment costs of the 3D printer and associated peripheral equipment notwithstanding). Taking into account the additional cost of the indispensable single-use pulmonary function filter for connection to the adapter (< \$ 1), the printed adapter costs around one-third to one-fifth of the price of the conventional facemask. Notably, the possibility to sterilise the adapters for reuse multiple times further considerably reduces its cost and thereby increases its cost efficiency in studies with large cohorts and/or longitudinal sampling over a broad timespan. (Sterilisation of the silicon facemask is limited due to its in-built sterility filter, which is susceptible to deterioration during autoclaving or chemical treatment).

Another limitation consideration of the facemask in relation to the ensuing breath sample analysis is that the material itself is a potential source of confounding compounds and might equally act as a sink for volatile breath constituents through adsorption. Although this cannot be ruled out for the 3D-printed sampling adapter presented here, at least one of the resins (BioMed Clear) is specified to exhibit no volatile emissions, so its potential as a source of confounders is low.

A last aspect of the adapter to consider is its resilience to the autoclaving (sterilisation) process, specifically the potential changes in the physical properties of the material induced by the treatment, e.g., micro-fractures or apparent porosity. Although no explicit material properties assessments were undertaken in the present work, visual inspection yielded no visible changes after autoclaving, apart from a slight shift in hue for the Surgical Guide resin models, changing from orange to pale-yellow. Further, its firm connection to the ReCIVA device, as well as the insertion of the TD tubes, remained viable after autoclaving, indicating that any dimensional changes were minimal and did not compromise its further use.

Finally, it is worth noting that the adapter design presented in this technical note is versatile in that it can be modified depending on the intended application and

equipment available for use as an adjunct to the ReCIVA device. As an example, the design can be adjusted to fit the requirements for sampling breath from children, or for use with an alternative/additional sterile filter located elsewhere within the sampling set-up. Notably, any modifications of a certified medical device conventionally render the certification invalid, thus this aspect must also be considered for the novel adapter presented here. However, the ReCIVA device is currently not certified as a medical device and is intended for research purposes only, thus the present modifications do not compromise this aspect of the sampler. Preliminary performance assessments indicate that exhaled breath sampled via the novel 3D-printed adapter achieves comparable levels of selected breath volatiles to samples collected with the commercial facemask [36]. Further, the ReCIVA system configuration presented in this technical note is currently in successful use in an ongoing clinical study, which reasserts its applicability in the intended setting [37].

Conclusions

3D printing in medical science is an emerging practice that offers manifold benefits, primarily the cost-efficient fabrication and design flexibility of bespoke parts for specific and/or niche applications. The field of breath analysis represents a novel approach to disease diagnostics in the clinical environment and beyond [21]. Limited access to suitable tools for sampling breath, for example, due to budget constraints in procuring commercial adapters, poses a hindrance to advancement in this field of research. Consequently, a low-cost 3D-printed sampling adapter, as presented in this technical note, represents a feasible alternative to commercial products. The flexibility and versatility of 3D printers provide a freedom to operate and produce custom-made medical products and parts [38]. The availability of easily-accessible designs, such as via open-access platforms, is essential for transparency of the model and incremental improvements proposed by the user community, as well as for subsequent proliferation for widespread and cost-effective implementation of the designed model. This is especially relevant in periods of emergency, when shortages in regular supply chains might cause limitations in medical care. Overall, the use of 3D printing is set to surpass its current emphasis on prototyping to become more widely adopted for commercial use, especially as this technology continues to mature and as prototype models are sufficiently tested for biocompatibility and safety for medical applications.

Abbreviations

CAD: Computer-aided design; ETO: ethylene oxide; GC-MS: Gas chromatography-mass spectrometry; IPA: Isopropyl alcohol (isopropanol); ReCIVA:

Respiration collector for in-vitro analysis; SLA: Stereolithography; STL: Standard tessellation language; TD: Thermal desorption; UV: Ultraviolet (irradiation).

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s41205-022-00150-y>.

Additional file 1: Figure S1. (left) Lateral view of the ReCIVA breath sampler equipped with a commercially available facemask. (right) Cross-sectional view of the silicon mask for an improved visibility of nose and mouth placement above the assembled thermal desorption tubes.

Acknowledgements

Manuel Krause (Fraunhofer IVV) and Fabian Müller (Fraunhofer ITEM) are acknowledged for their technical support in modifying the design and printing the mouthpiece adapters. Christian Zacherl (Fraunhofer IVV) is thanked for configuring and photographing the models depicted in the figures. The article processing charge for this publication has been funded by an unrestricted grant from Formlabs.

Disclaimer

During the preparation of this manuscript, an alternative sampling interface for the ReCIVA device was launched by the manufacturer, Owlstone Medical Ltd. (Cambridge, UK) (<https://www.owlstonemedical.com/products/reciva/>). The new adapter has similarities with the design presented here, yet differs in terms of the materials used and the positioning of the sterile filter, which remains located downstream of the TD tubes. It is beyond the scope of this technical note to appraise the pros and cons of this new commercial design in comparison with the 3D-printed design reported here, yet certainly many of the disadvantages associated with the silicon facemask have been overcome with this new commercial adapter. Nevertheless, the possibility for self-printing and adapting the present design as a cost-effective alternative remains warranted.

Authors' contributions

OH designed and developed the prototype of the adapter. The initial 3D version was created and designed by OH, AC and FW. AC and FW established different techniques to digitally design variant 3D models. YP and JB tested and gave feedback for optimisation of the design of the adapter model and made contributions to the final version of the adapter described in this manuscript. YP drafted the manuscript. JB co-wrote and critically revised the manuscript. All authors read and approved the final manuscript.

Funding

Open Access funding enabled and organised by Projekt DEAL. This work was partly funded by the German Ministry of Education and Research (BMBF), grant number 01KT1803A, as part of the ERA-NET TRANSCAN-2 project Airborne Biomarkers for Colorectal Cancer (ABC-Cancer), as well as by the German Centre for Lung Research (DZL).

Availability of data and materials

All data and 3D printer files used in this technical note are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Competing interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author details

¹Fraunhofer Institute for Process Engineering and Packaging IVV, Giggenhauser Straße 35, 85354 Freising, Germany. ²Department of Chemistry and Pharmacy, Chair of Aroma and Smell Research, Friedrich-Alexander-Universität Erlangen-Nürnberg, Henkestraße 9, 91054 Erlangen, Germany. ³Fraunhofer Institute for Toxicology and Experimental Medicine ITEM, Feodor-Lynen-Str. 15, 30625 Hannover, Germany. ⁴Member of the German Centre of Lung Research DZL (BREATH), Hannover, Germany.

Received: 24 February 2022 Accepted: 7 June 2022

Published online: 09 August 2022

References

- Capobussi M, Moja L. An open-access and inexpensive 3D printed otoscope for low-resource settings and health crises. *3D Print Med.* 2021;7(1):1–8.
- Hagen A, Chisling M, House K, Katz T, Abelseth L, Fraser I, et al. 3D printing for medical applications: Current state of the art and perspectives during the COVID-19 crisis. *Surgeries.* 2021;2(3):244–59.
- Schubert C, Van Langeveld MC, Donoso LA. Innovations in 3D printing: a 3D overview from optics to organs. *Br J Ophthalmol.* 2014;98(2):159–61.
- Cavallo L, Marciandò A, Cicciù M, Oteri G. 3D printing beyond dentistry during COVID 19 epidemic: a technical note for producing connectors to breathing devices. *Prosthesis.* 2020;2(2):46–52.
- Oladapo BI, Ismail SO, Afolalu TD, Olowade DB, Zahedi M. Review on 3D printing: Fight against COVID-19. *Mater Chem Phys.* 2021;258:123943.
- Erickson MM, Richardson ES, Hernandez NM, Bobbert DW II, Gall K, Fearis P. Helmet modification to PPE with 3D printing during the COVID-19 pandemic at Duke University Medical Center: a novel technique. *J Arthroplasty.* 2020;35(7):S23–S7.
- Radfar P, Bazaz SR, Mirakhorli F, Warkiani ME. The role of 3D printing in the fight against COVID-19 outbreak. *J 3D Print Med.* 2021;5(1):51–60.
- Koprnický J, Najman P, Šafka J. 3D printed bionic prosthetic hands. 2017 IEEE International Workshop of Electronics, Control, Measurement, Signals and their Application to Mechatronics (ECMSM); 2017.
- Mee H, Greasley S, Whiting G, Harkin C, Oliver G, Marsden D, et al. 3D printed customised external cranial plate in a patient with syndrome of trephined: 'a case report'. *3D Print Med.* 2021;7(1):35.
- Faldini C, Mazzotti A, Belvedere C, Durastanti G, Panciera A, Geraci G, et al. A new ligament-compatible patient-specific 3D-printed implant and instrumentation for total ankle arthroplasty: from biomechanical studies to clinical cases. *J Orthop Traumatol.* 2020;21(1):1–9.
- Jammalamadaka U, Tappa K. Recent advances in biomaterials for 3D printing and tissue engineering. *J Funct Biomater.* 2018;9(1):22.
- Mironov V, Kasyanov V, Markwald RR. Organ printing: from bioprinter to organ biofabrication line. *Current Opinion in Biotechnology.* 2011;22(5):667–73.
- Hopfner C, Jakob A, Tengler A, Grab M, Thierfelder N, Brunner B, et al. Design and 3D printing of variant pediatric heart models for training based on a single patient scan. *3D Print Med.* 2021;7(1):1–11.
- Gillett D, Bashari W, Senanayake R, Marsden D, Koulouri O, MacFarlane J, et al. Methods of 3D printing models of pituitary tumors. *3D Print Med.* 2021;7(1):24.
- Wake N, Rosenkrantz AB, Huang WC, Wysock JS, Taneja SS, Sodickson DK, et al. A workflow to generate patient-specific three-dimensional augmented reality models from medical imaging data and example applications in urologic oncology. *3D Printing in Medicine.* 2021;7(1):34.
- Leukers B, Gülkan H, Irsen SH, Milz S, Tille C, Schieker M, et al. Hydroxyapatite scaffolds for bone tissue engineering made by 3D printing. *J Mater Sci: Mater Med.* 2005;16(12):1121–4.
- Mironov V, Boland T, Trusk T, Forgacs G, Markwald RR. Organ printing: computer-aided jet-based 3D tissue engineering. *Trends Biotechnol.* 2003;21(4):157–61.
- Ventola CL. Medical applications for 3D printing: current and projected uses. *Pharm Ther.* 2014;39(10):704.
- Pleil JD, Stiegel MA, Beauchamp JD. Chapter 1 - Breath biomarkers and the exposome. In: Beauchamp J, Davis C, Pleil J, editors. *Breathborne Biomarkers and the Human Volatilome*. 2nd ed. Amsterdam: Elsevier; 2020. p. 3–21.

20. Drabińska N, Flynn C, Ratcliffe N, Belluomo I, Myridakis A, Gould O, et al. A literature survey of all volatiles from healthy human breath and bodily fluids: the human volatilome. *J Breath Res.* 2021;15(3):034001.
21. Pham YL, Beauchamp J. Breath Biomarkers in Diagnostic Applications. *Molecules.* 2021;26(18):5514.
22. Davis MD, Fowler SJ, Montpetit AJ. Exhaled breath testing – A tool for the clinician and researcher. *Paediatr Respir Rev.* 2019;29:37–41.
23. Bruderer T, Gaisl T, Gaugg MT, Nowak N, Streckenbach B, Müller S, et al. On-Line Analysis of Exhaled Breath: Focus Review. *Chem Rev.* 2019;119(19):10803–28.
24. Pereira J, Porto-Figueira P, Cavaco C, Taunk K, Rapole S, Dhakne R, et al. Breath analysis as a potential and non-invasive frontier in disease diagnosis: an overview. *Metabolites.* 2015;5(1):3–55.
25. Beauchamp JD, Miekisch W. Chapter 2 - Breath sampling and standardization. In: Beauchamp J, Davis C, Pleil J, editors. *Breathborne Biomarkers and the Human Volatilome.* 2nd ed. Amsterdam: Elsevier; 2020. p. 23–41.
26. Herbig J, Beauchamp J. Towards standardization in the analysis of breath gas volatiles. *J Breath Res.* 2014;8(3):037101.
27. Harshman SW, Pitsch RL, Davidson CN, Lee EM, Scott AM, Hill EM, et al. Evaluation of a standardized collection device for exhaled breath sampling onto thermal desorption tubes. *J Breath Res.* 2020;14(3):036004.
28. Khan MF, Sahani M, Nadzir MSM, Yik LC, Hoque HMS, Abd Hamid HH, et al. Volatile organic compound analysis by sorbent tube-thermal desorption-gas chromatography: A review. *Int J Eng Technol (UAE).* 2018;7(3.14 Special Issue 14):165–75.
29. Harshman SW, Pitsch RL, Davidson CN, Scott AM, Hill EM, Smith ZK, et al. Characterization of standardized breath sampling for off-line field use. *J Breath Res.* 2019;14(1):016009.
30. Doran SLF, Romano A, Hanna GB. Optimisation of sampling parameters for standardised exhaled breath sampling. *J Breath Res.* 2017;12(1):016007.
31. Holden KA, Ibrahim W, Salman D, Cordell R, McNally T, Patel B, et al. Use of the ReCIVA device in breath sampling of patients with acute breathlessness: a feasibility study. *ERJ Open Res.* 2020;6(4):00119–2020.
32. Khan MS, Cuda S, Karere GM, Cox LA, Bishop AC. Breath biomarkers of insulin resistance in pre-diabetic Hispanic adolescents with obesity. *Sci Rep.* 2022;12(1):339.
33. Wang T, Pysanenko A, Dryahina K, Španěl P, Smith D. Analysis of breath, exhaled via the mouth and nose, and the air in the oral cavity. *J Breath Res.* 2008;2(3):037013.
34. Chen W, Metsälä M, Vaittinen O, Halonen L. The origin of mouth-exhaled ammonia. *J Breath Res.* 2014;8(3):036003.
35. Chen W, Metsälä M, Vaittinen O, Halonen L. Hydrogen cyanide in the headspace of oral fluid and in mouth-exhaled breath. *J Breath Res.* 2014;8(2):027108.
36. Holz O, Boehm C, Behrendt M, Guenther F, Schuchardt S, Hohlfeld JM, editors. Testing and customization of the ReCIVA breath sampler. Poster presented at: IABR Breath Summit 2019; 2019 08.-11.09.2022; Loughborough, UK.
37. Kenn K, Gloeckl R, Leitl D, Schneeberger T, Jarosch I, Hitzl W, et al. Protocol for an observational study to identify potential predictors of an acute exacerbation in patients with chronic obstructive pulmonary disease (the PACE Study). *BMJ Open.* 2021;11(2):e043014.
38. Banks J. Adding value in additive manufacturing: researchers in the United Kingdom and Europe look to 3D printing for customization. *IEEE Pulse.* 2013;4(6):22–6.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

